Biosecurity Imperatives and the Feed Supply Chain



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Why biosecurity?

biosecurity noun

Save Word

bio·se·cu·ri·ty | \ bī-(,)ō-si-ˈkyür-ə-tē 🕥 \

Definition of *biosecurity*

: security from exposure to harmful biological agents *also* : measures taken to ensure this security

Merriam-Webster



- Bi-directional:
 - Bio-exclusion keeping pathogens out
 - Bio-containment preventing pathogens from spreading



Biosecurity

- Practices of biosecurity have been widely implemented in the swine industry
 - Entry benches
 - Shoe covers
 - Downtime
 - Shower in/shower out
 - Disinfection of incoming supplies









Pathogens capable of surviving in feed and/or feed ingredients

- Bacteria
 - Salmonella spp.
 - Escherichia coli
- Viruses
 - Porcine epidemic diarrhea virus (PEDV)
 - African swine fever virus (ASFV)
 - Senecavirus A (SVA)
 - Classical swine fever virus (CSF)
 - Pseudorabies virus (PRV)
 - Foot and mouth disease (FMD)
- Others?

Survival of viral pathogens in animal feed ingredients under transboundary shipping models

Scott A. Dee¹*, Fernando V. Bauermann², Megan C. Niederwerder^{3,4}, Aaron Singrey², Travis Clement², Marcelo de Lima^{2,5}, Craig Long², Gilbert Patterson⁶, Maureen A. Sheahan³, Ana M. M. Stoian³, Vlad Petrovan³, Cassandra K. Jones⁷, Jon De Jong¹, Ju Ji⁸, Gordon D. Spronk¹, Luke Minion¹, Jane Christopher-Hennings², Jeff J. Zimmerman⁹, Raymond R. R. Rowland³, Eric Nelson², Paul Sundberg¹⁰, Diego G. Diel²

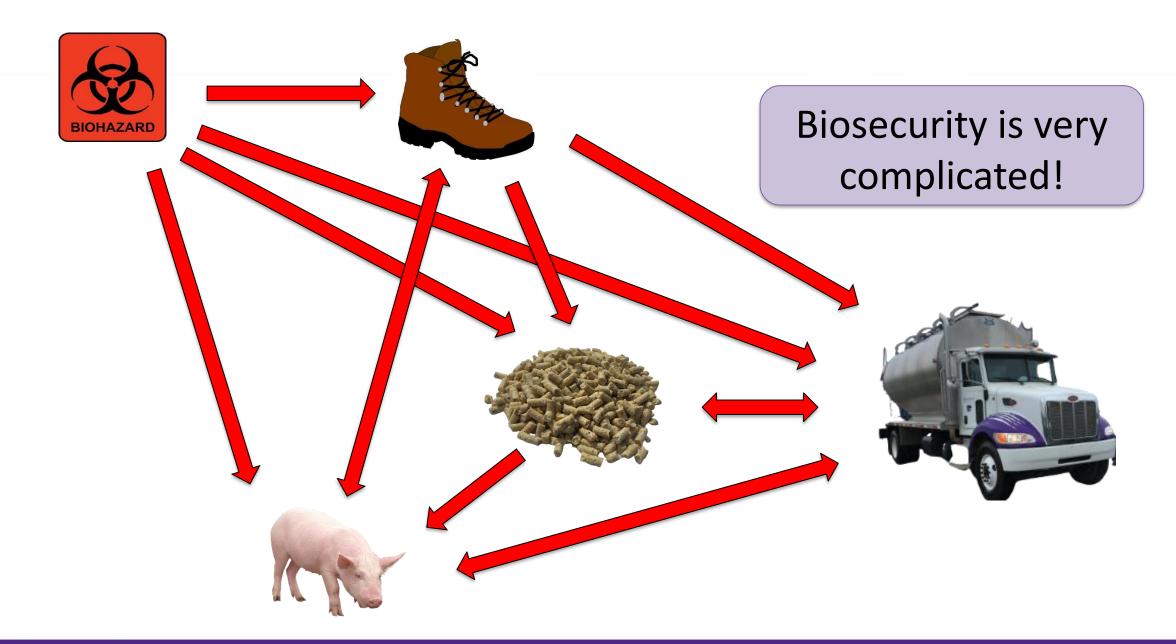
Evaluation of the minimum infectious dose of porcine epidemic diarrhea virus in virus-inoculated feed

OBJECTIVE Loni L. Schumacher DVM To determine the minimum infectious dose of porcine epidemic diarrhe lason C. Woodworth PhD virus (PEDV) in virus-inoculated feed. Cassandra K. Jones PhD Oi Chen MS. DVM ANIMALS Jianqiang Zhang PhD 30 crossbred 10-day-old pig Phillip C. Gauger DVM, PhD PROCEDURES Charles R. Stark PhD Tissue culture PEDV was diluted to form 8 serial 10-fold dilutions. An ali-quot of stock virus (5.6 X 10⁵ TCID₆₀/mL) and each serial PEDV dilution Rodger G. Main DVM, PhD Richard A. Hesse PhD were mixed into 4.5-kg batches of feed to create 9 PEDV-inoculated feed doses; I virus-negative dose of culture medium in feed was also created Mike D. Tokach PhD Pigs were challenge exposed via oral administration of PEDV-inoculate Steve S. Dritz DVM, PhD feed, and fecal swab specimens were collected. All pigs were euthanized 7 days after challenge exposure; fresh tissues were collected and used for PCR assay, histologic examination, and immunohistochemical analysis. Received November 18, 2015. Accepted December 21, 2015. From the Department of Diagnostic Medicinel/Patho-biology, College of Veterinary Medicine (Schumacher, Hease, Dritz), and the Department of Animal Schu-Steiner and Industry (Jones, Sark), College of Ag-riculture, Kanas State, University, Mahattan, KS 66506; and the Department of Veterinary Diagnostic and Production Animal Medicine, College of Veteri-nary Medicine, Iowa State University, Ameta, IA 50011 (Chen, Zhang, Cauger, Huin). RESULTS The PCR cycle threshold (Ct) decreased by approximately 10 when PEDV was added to feed, compared with results for equivalent PEDV diluted in tissue culture medium. Pigs became infected with PEDV when challenge exposed with the 4 highest concentrations (lowest concentration to cause infection, 5.6 \times 10¹ TCID₅₀/g; Ct = 27 in tissue culture medium and 37 ii CONCLUSIONS AND CLINICAL RELEVANCE In this study, PEDV in feed with detectable Ct values of 27 to 37 was in-fective. The Ct was 37 for the lowest infective PEDV dose in feed, which Address correspondence to Dr. Schumacher (Ionil@ may be above the limit of detection established for PEDV PCR assays used by some diagnostic laboratories. Overall, results indicated 5.6 X 10 TCID₅₀/g was the minimum PEDV dose in feed that can lead to infec tion in 10-day-old pigs under the conditions of this study. (Am I Vet Re 2016:77:1108-1113

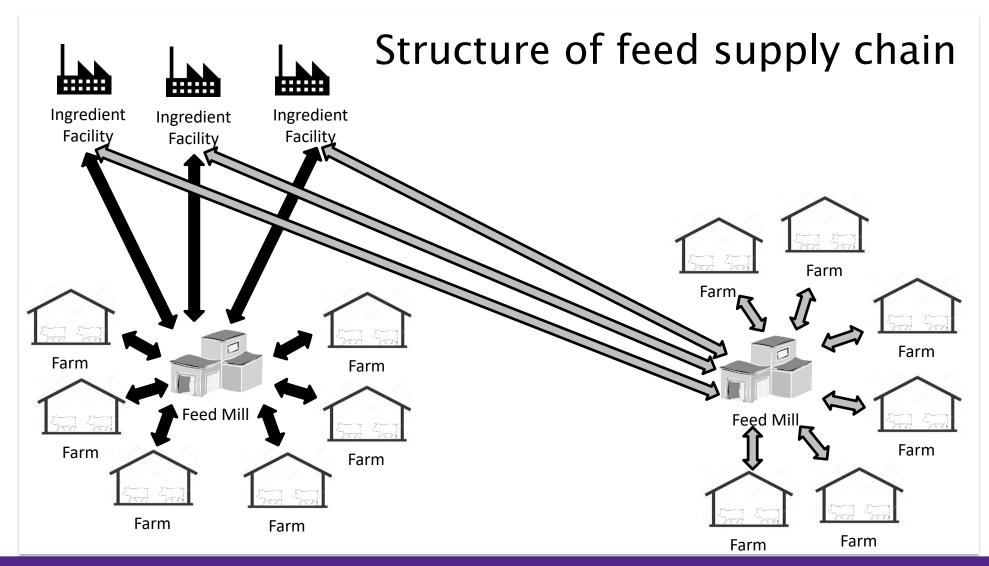
Infectious Dose of African Swine Fever Virus When Consumed Naturally in Liquid or Feed

Megan C. Niederwerder, Ana M.M. Stoian, Raymond R.R. Rowland, Steve S. Dritz, Vlad Petrovan, Laura A. Constance, Jordan T. Gebhardt, Matthew Olcha, Cassandra K. Jones, Jason C. Woodworth, Ying Fang, Jia Liang, Trevor J. Hefley

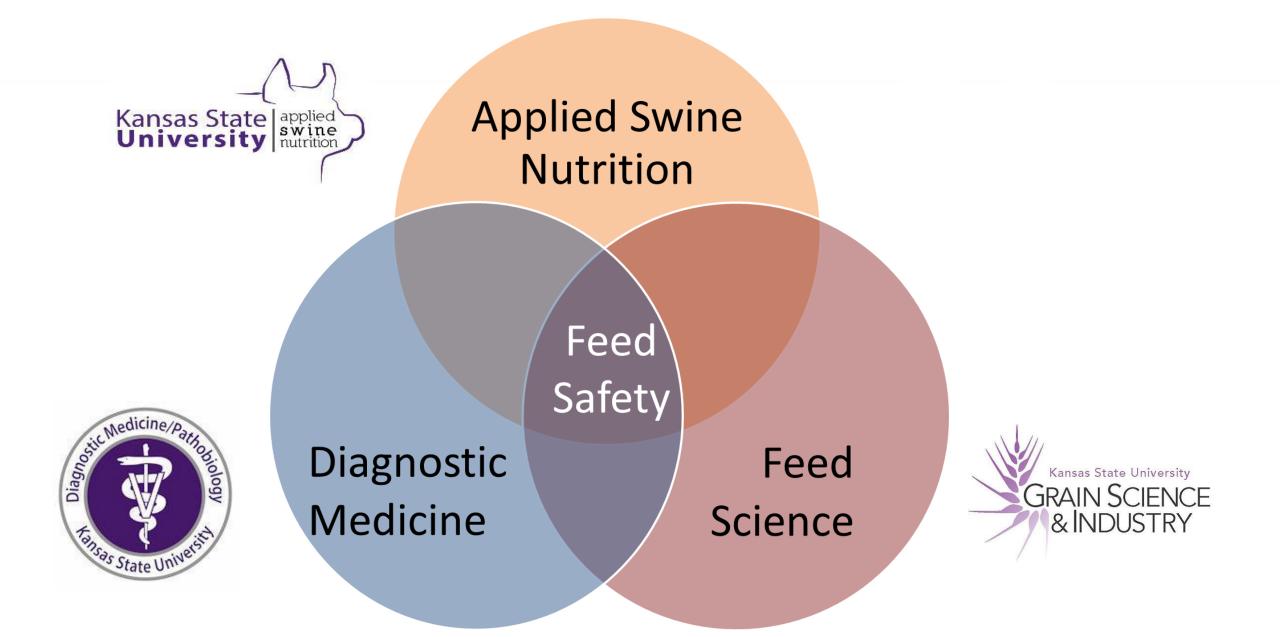






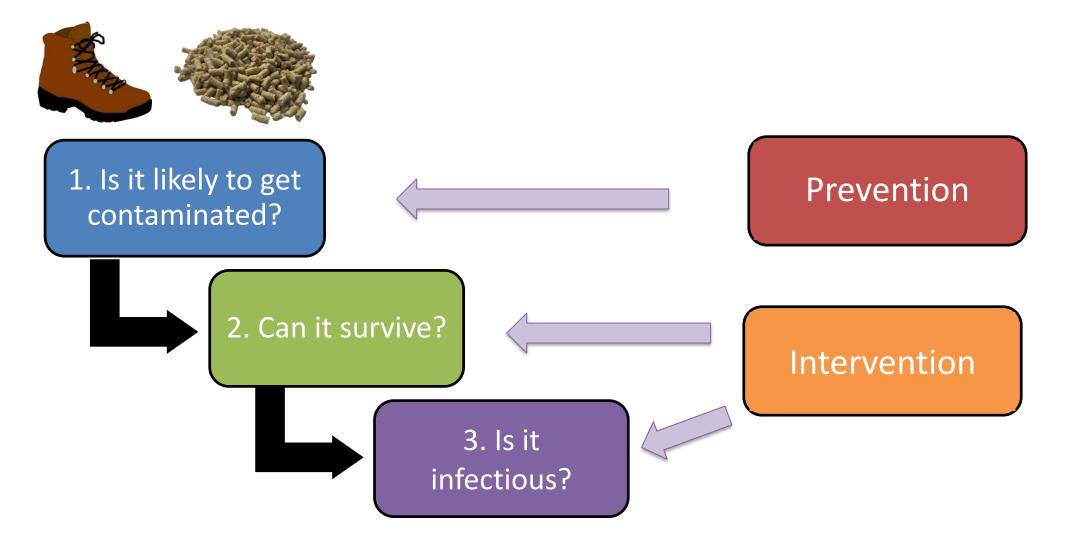








Addressing Feed Safety





- Risk of contamination depends on:
 - Geographical considerations
 - Countries/regions with active disease outbreaks
 - Location of pigs with disease relative to location of ingredient production
 - Agricultural practices
 - Packaging
 - Single use bags or totes vs. re-used totes or bulk trailers







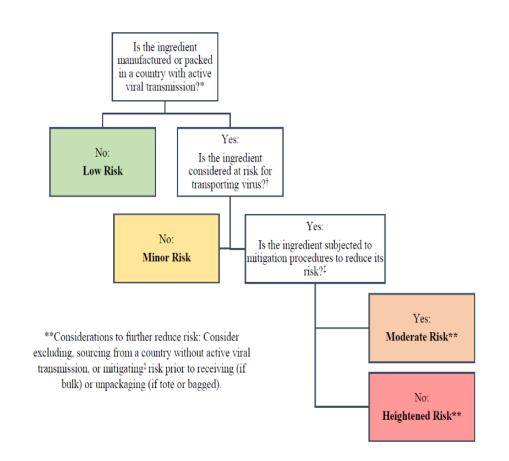




- Understand where ingredients are coming from
- Are alternative sources available and cost effective?
- BIOSECURITY during manufacture, storage, and delivery









https://www.swinehealth.org/wp-content/uploads/2018/09/Feed-Ingredient-Safety.pdf



Areas where improvement would be beneficial













Research partnership

Production system located in Vietnam

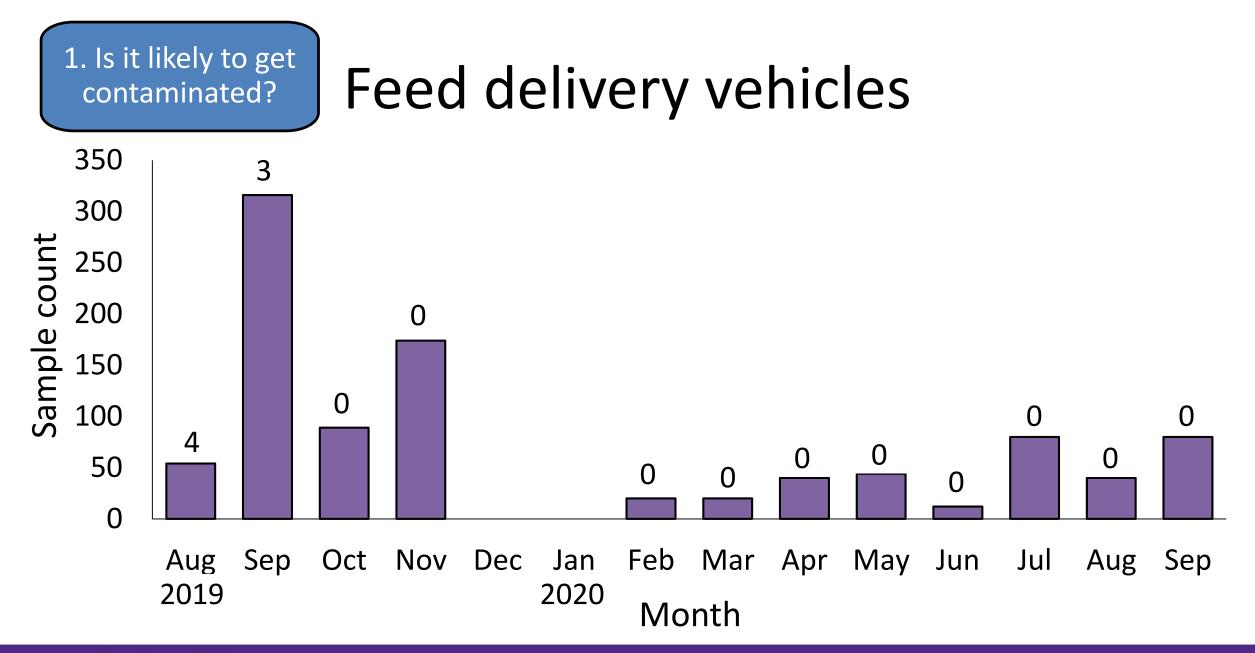
Goal: Use diagnostic testing capabilities to understand the risk of ASFV spread within their production system

- 1. Feed production system
 - a. Feed mill
 - b. Ingredients and finished feed
 - c. Feed trucks
- 2. Live animal transport
- 3. Market animal transfer center

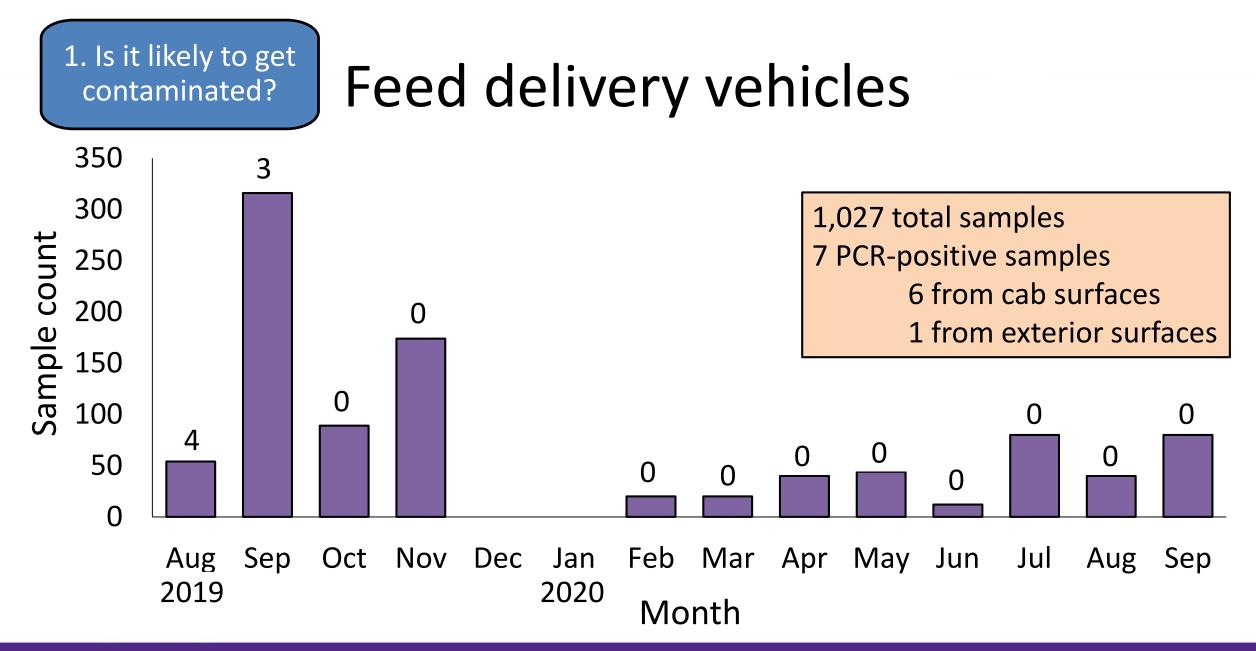














How can this be accomplished?







Step 1: Remove organic material

Step 2: Dry Step 3: Apply disinfectant



How can this be accomplished?

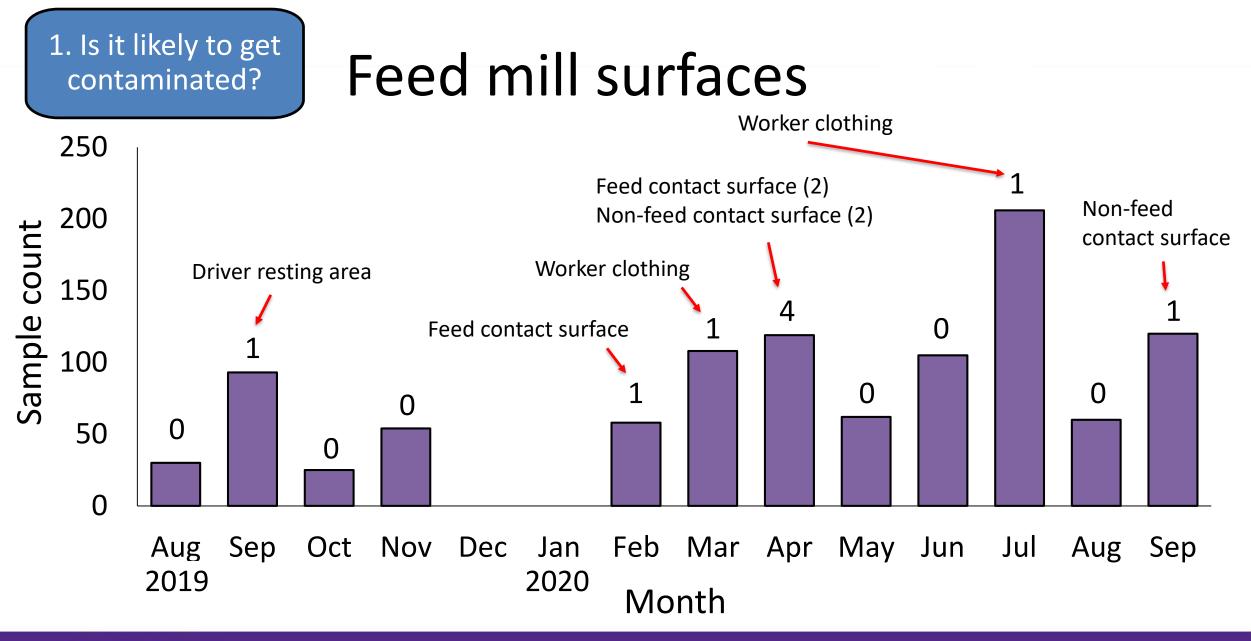


Avoid this

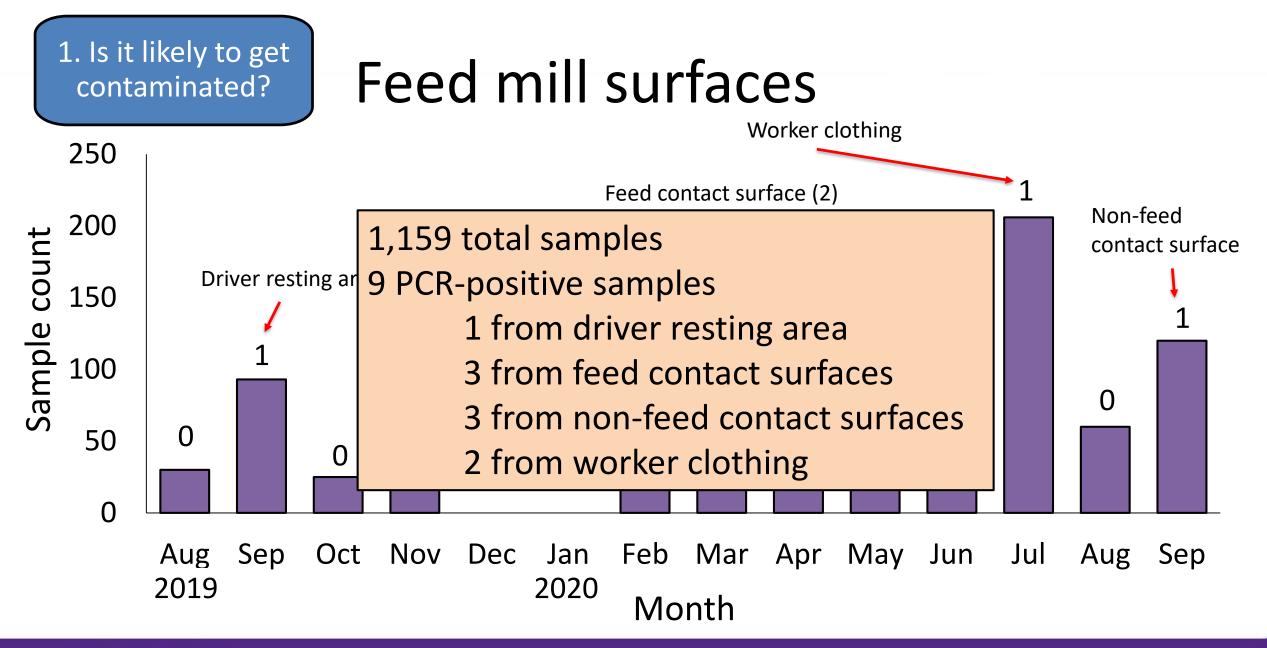








K-STATE Research and Extension



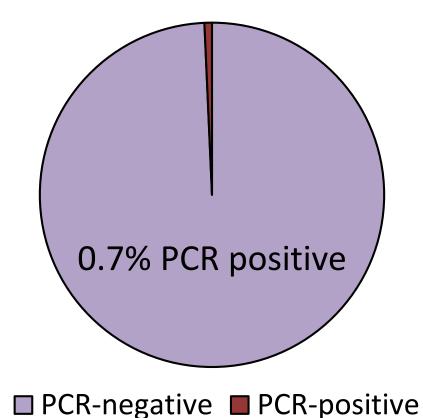


Feed and ingredients

142 total samples so far

40 ingredient and water samples 102 complete feed samples

- 1 complete feed sample PCR positive
- Batch of feed did not contain added formaldehyde-based product

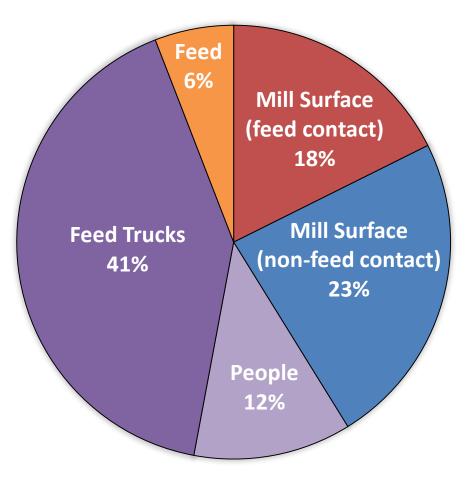


Research and Extension

Where is the contamination at?

- 17 of 2,328 samples (0.7%) from the feed supply chain contain ASFV DNA as determined by PCR
 - \circ 3 Feed-Contact Surfaces in Mill
 - $\circ~$ 4 Non-Feed-Contact Surfaces in Mill
 - 2 Employee clothing in Mill
 - \circ 1 Complete Feed
 - 7 Feed Trucks

<u>Key finding:</u> People and fomites are incredibly important!





Risk of ASFV carryover if feed after contaminated batch



1

2

3

4

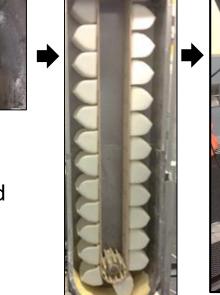
5

6





Batch Ingredients Negative **ASFV** Inoculated Negative Negative Negative Negative











Risk of ASFV carryover if <u>feed</u> after contaminated batch

Detection of African swine fever virus (ASFV) p72 DNA in feed samples					
		Batch of feed			
	1				
Batch	Negative				
Non-detected	10				
Suspect	0				
ASFV detected	0				





Risk of ASFV carryover if <u>feed</u> after contaminated batch

Detection of African swine fever virus (ASFV) p72 DNA in feed samples							
	Batch of feed						
	1 2						
Batch	Negative	Positive					
Non-detected	10	0					
Suspect	0	0	-				
ASFV detected	0	10	-				





Risk of ASFV carryover if <u>feed</u> after contaminated batch

Detection of African swine fever virus (ASFV) p72 DNA in feed samples									
		Batch of feed							
	1	1 2 3							
Batch	Negative	Positive							
Non-detected	10	0	0						
Suspect	0	0	0						
ASFV detected	0	10	10						





Risk of ASFV carryover if <u>feed</u> after contaminated batch

Detection of Afric	Detection of African swine fever virus (ASFV) p72 DNA in feed sa								
			Batch	of feed					
	1	1 2 3 4							
Batch	Negative	Negative Positive Negative Negative							
Non-detected	10	0	0	0					
Suspect	0	0	0	1					
ASFV detected	0	10	10	9					





Risk of ASFV carryover if <u>feed</u> after contaminated batch

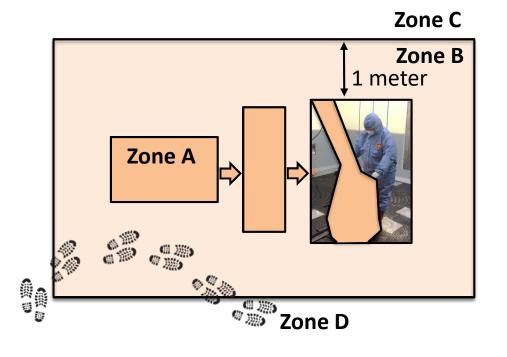
Detection of African swine fever virus (ASFV) p72 DNA in feed samples									
	Batch of feed								
	1	1 2 3 4 5 6							
Batch	Negative	Positive	Negative	Negative	Negative	Negative			
Non-detected	10	0	0	0	0	0			
Suspect	0	0	0	1	1	3			
ASFV detected	0	10	10	9	9	7			



ASFV was still detected after 4 subsequent batches of feed



Risk of ASFV carryover on <u>feed surfaces</u> <u>and within environment</u> after contaminated batch

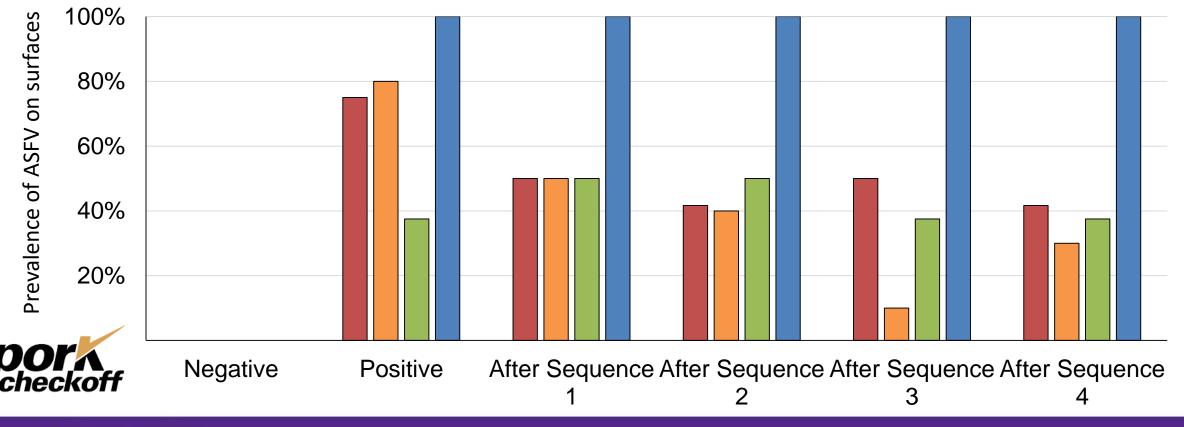






Risk of ASFV carryover on <u>feed surfaces</u> <u>and within environment</u> after contaminated batch

■ Feed Contact Surface ■ < 1 m ■ > 1 m ■ Shoes





Elijah et al., 2021

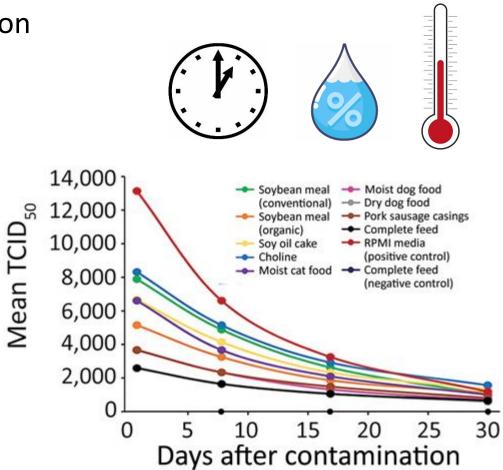
Risk of ASFV carryover on <u>feed surfaces</u> <u>and within environment</u> after contaminated batch

- Key findings:
 - ASFV has similar characteristics to PEDV within a feed mill
 - It goes everywhere!
 - Contamination of feed and surfaces can be detected after multiple batches of feed pass through the equipment
 - People are extremely important to consider!



2. Can it survive

- Pathogen has to survive on surface to cause infection
 - Viruses do not replicate outside of host
 - Naturally decay over time (lose infectivity)
 - Time, temperature, humidity, environment
- Greatest survival in:
 - \circ Choline
 - Soybean meal
 - \circ Soy oil cake

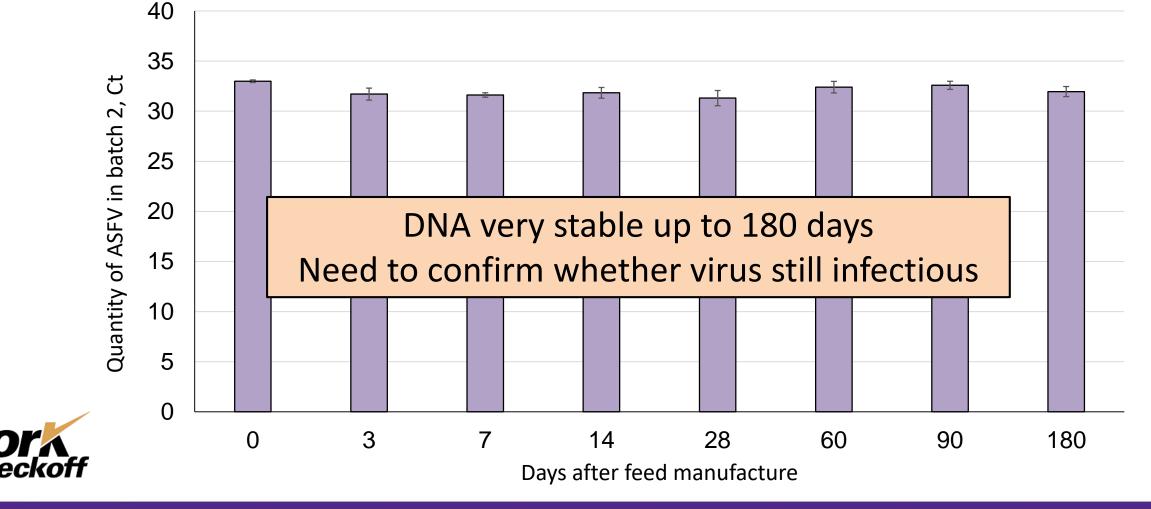




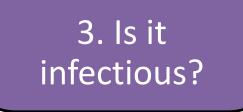
Stoian et al., 2019

2. Can it survive

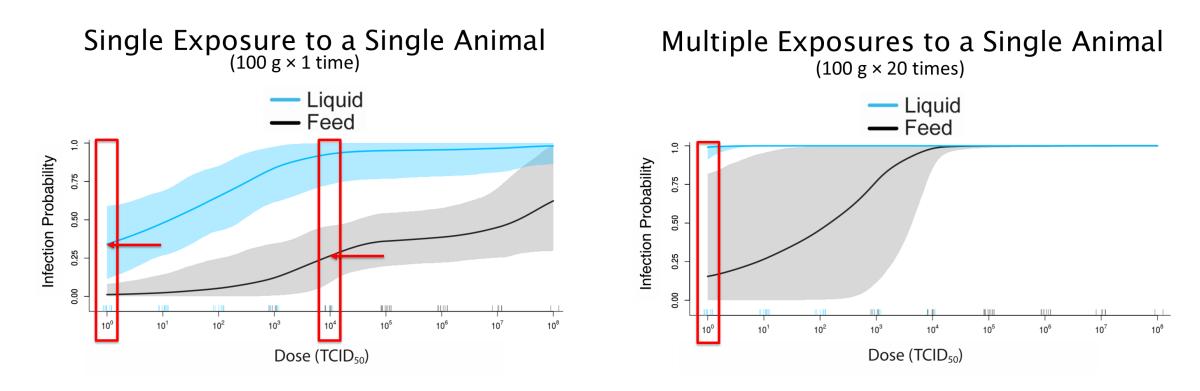
Detection of African swine fever virus in contaminated <u>feed</u> over time







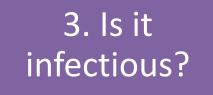
Feed as vector for disease



Multiple exposures increases risk of infection



Niederwerder et al., 2019



Feed as vector for disease

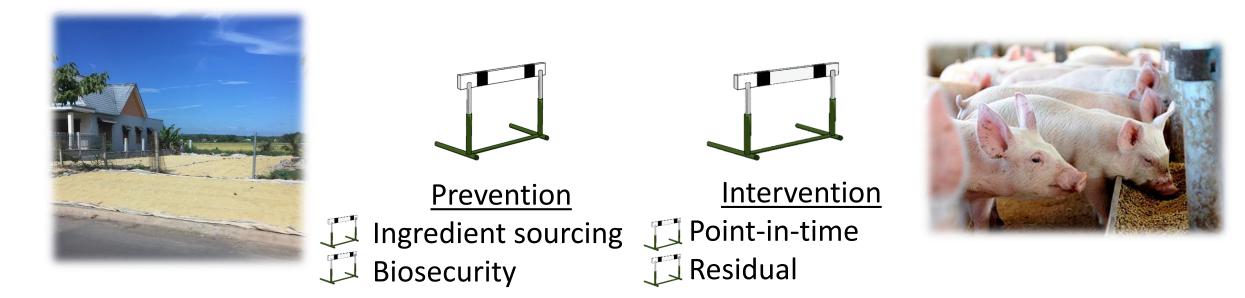
 For PEDV, 1 gram of feces from an acutely infected pig can contaminate 500 tonnes of feed – with EACH GRAM being infective





Schumacher et al., 2016

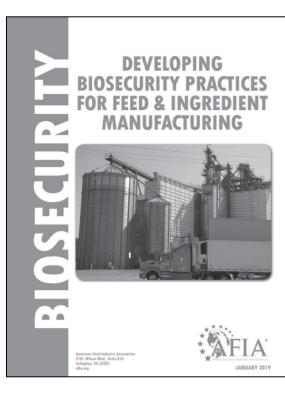
Feed Biosecurity: Hurdles to Prevent Pathogen Transfer through feed supply chain





Prevention

• Biosecurity at feed mills



Feed mill biosecurity plans: A systematic approach to prevent biological pathogens in swine feed

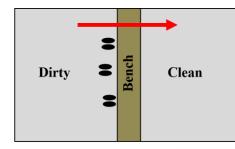
Roger A. Cochrane, MS; Steve S. Dritz, DVM, PhD; Jason C. Woodworth, MS, PhD; Charles R. Stark, MS, PhD; Anne R. Huss, MS, PhD; Jean Paul Cano, DVM, PhD; Robert W. Thompson, DVM, MS; Adam C. Fahrenholz, MS, PhD; Cassandra K. Jones, MS, PhD





Prevention

- Extend biosecurity to feed mills to limit contamination from trucks and people
 - Use receiving mats/funnels
 - When possible, don't let drivers out of trucks
 - Use your own employees to unload
 - Start treating your mill like your farm physical barriers, foot baths, zoning
- Consider truck disinfection





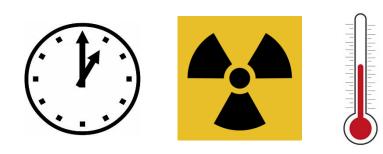




Point-in-Time

<u>Residual</u>

- Susceptible to recontamination Have some level of residual activity
 - Time
 - Irradiation
 - Thermal processing



to help combat possible recontamination

- Acids and alkalis
- Essential oils
- Formaldehyde-based products
- Medium chain fatty acids



Intervention

Point in time: Holding time

- Based on half-life estimates, recommended holding times have been established
 Mean Holding Time for 99.99% SVA Degradation
 - 1. Temperature
 - 2. Humidity
 - 3. Ingredient matrix

Mean Holding Time for 99.99% SVA Degradation								
Days at Days at Days at 4°C (39.6°F) 15°C (59°F) 30°C (86°F)								
Conventional SBM	143 days	52 days	26 days					
DDGS	494 days	182 days	26 days					
Vitamin D	39 days	26 days	26 days					
Lysine	78 days	13 days	13 days					

Mean Holding Time for 99.99% ASF Degradation at 54°F Avg.³



mean norung time for 55.55% AST Degradation at 54 T Avg.							
	Average	95% Confidence Interval - Lower	95% Confidence Interval – Higher				
Conventional SBM	125 days	113 days	135 days				
Organic SBM	168 days	150 days	186 days				
Choline	155 days	142 days	168 days				



https://www.swinehealth.org/wp-content/uploads/2020/02/Holding-Time-Calculations-for-Feed-Ingredients-to-Mitigate-Virus-Transmission-Print-02.04.20.pdf

Intervention

Point in time: Thermal processing

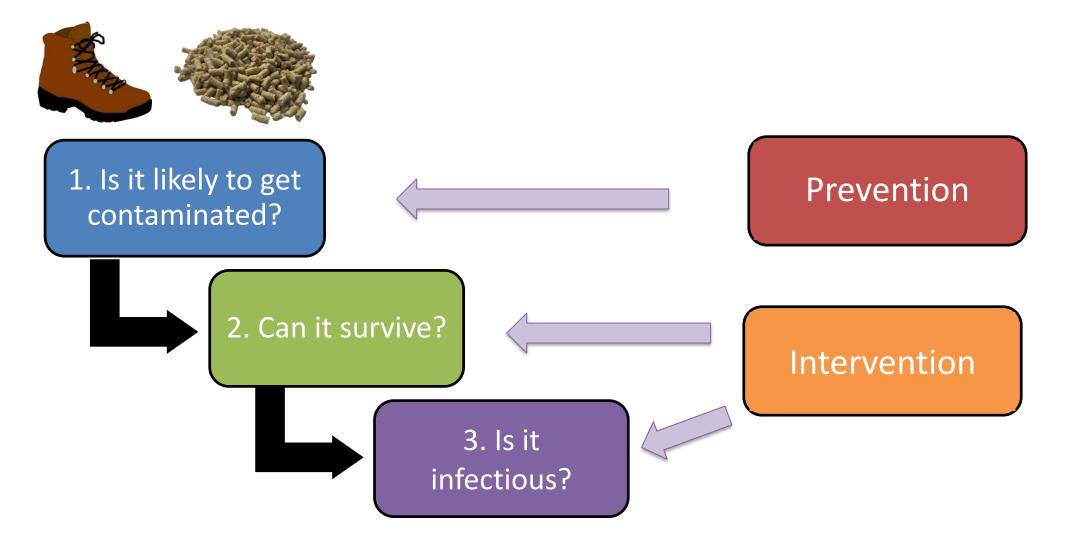
	Feed	0 dpi	2 dpi	4 dpi	6 dpi	7 dpi	7 dpi Cecum
No PEDV	0	0	0	0	0	0	0
38°C	9/9	0	1/9	3/9	3/9	3/9	3/9
46°C	9/9	0	3/9	3/9	3/9	3/9	3/9
54°C	9/9	0	0	0	0	0	0
63°C	8/9	0	0	0	0	0	0
71°C	8/9	0	0	0	0	0	0

No infectivity in diets pelleted \geq 54°C (129°F)



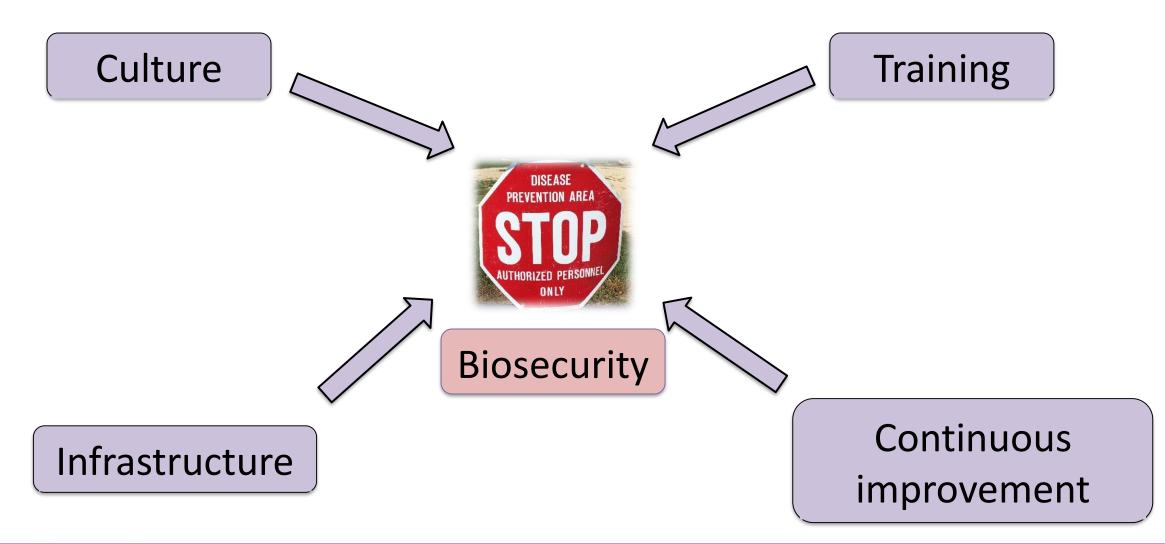
Cochrane et al., 2017

Addressing Feed Safety





Keys to a successful biosecurity program







- Any biosecurity program cannot be successful without the proper culture
 - Support and feedback from management
 - Financial support
- Consistent expectations and accountability at all levels of organization





- SOP's are worthless if employees don't know how to implement
- Routine reinforcement
- Focus on the <u>WHY</u>
- Glo-Germ fluorescent powder



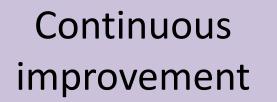


Infrastructure

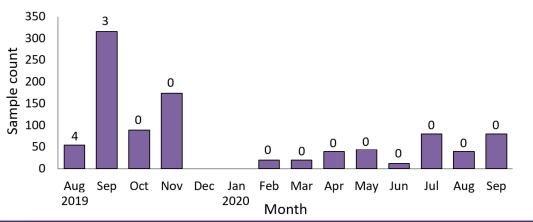
- Facility designs must accommodate biosecurity practices
 - Minimize inconvenience = more consistent implementation

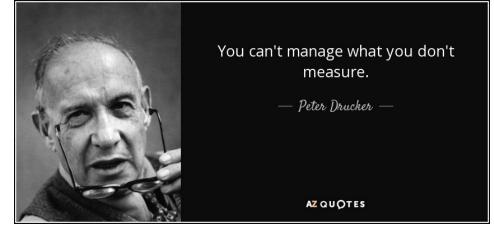






- Routine biosecurity audits of mills and mills
 - Change the paradigm and negative perception
 - Getting better helps everybody
- Sampling and diagnostic testing

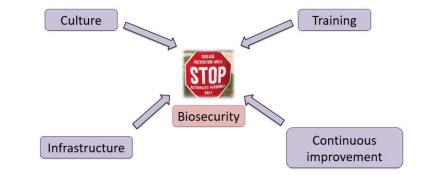






What have we learned

- Biosecurity is:
 - Inconvenient
 - Expensive



- Modern swine production is continuously moving towards high health
- Biosecurity as a whole is critical to long-term success of swine businesses
 - Feed biosecurity is becoming a critical component



<u>Kansas State University Feed Safety Team</u> Dr. Jordan Gebhardt – Diagnostic Medicine/Pathobiology

- Dr. Cassie Jones Animal Sciences & Industry
- Dr. Chad Paulk Feed Science

Dr. Jason Woodworth – Animal Sciences & Industry

<u>www.ksuswine.org</u> \rightarrow Feed Safety Resources

